A Survey on Selected Quality Parameters of Buffalo Milk Samples Collected from Consumer Markets of Three Different Central Governorates in Egypt

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Abstract
One hundred and twenty samples of raw buffalo milk were collected at consumer markets in central Cairo, Giza and Qualubya, Egypt. All samples were analysed chemically using Lactoscan, and microbiologically using the pour plate technique. Also, levels of aflatoxin (AF) M1 were assessed by using a commercial ELISA kit. The mean values were -0.476±0.07 °C, 5.89±0.79 %, 4.13±0.53 %, 4.25±0.53 %, 0.69±0.14 %, and 14.59±1.39 % for freezing point, fat, protein, lactose, ash and total solids respectively. Bacteriological enumeration of total mesophilic aerobic bacteria, coliform, spore forming bacteria and psychrotrophic bacteria were 5.37±0.78, 3.77±0.90, 2.78±0.36 and 2.84±0.39 Log CFU/ml respectively. The median concentration of AF M1 in all samples was 44.31 ng/L. Our results indicate that there is an essential need for improving the hygienic conditions in the production of raw milk.

Key Words: Milk quality, safety, aflatoxin M1, codex, HACCP, ISO 22000

Introduction
Nutritionally, milk has been defined as “the most nearly perfect food”, it is part of the daily diet and is important for growing children [1]. Moreover, the consumer demand for high quality milk and dairy products has a significant impact on all aspects of the dairy chain [2]. Buffalo milk receives an increasing research interest and investment in various countries because of its higher nutrient content compared to cow milk [3,4].

Freezing point, chemical composition (mainly fat, protein, lactose and total solids) and microbiological analysis of raw milk samples are important indicators of its quality. Measurement of the freezing point is used to detect any milk adulteration with water [5]. In Egypt, the average of the freezing point of buffalo milk samples ranges from -0.552 to -0.558 °C [6] and the regulatory standard ranges from -0.530 to -0.560 °C [7]. A freezing point of -0.528 °C for raw buffalo milk has been found in Italy [8] and Poland [9].

As with any type of raw milk, different microbial groups (lactic acid bacteria, coliform, Staphylococcus spp., Listeria spp., yeast and moulds, etc.) can rapidly multiply because of its high nutrient content. At cold storage temperatures, an increase in growth and metabolic activities of psychrotrophic bacteria can be noticed. Some of them are able to produce heat resistant proteolytic and lipolytic enzymes that can survive during UHT processing [10-12] resulting in the deterioration of UHT milk.

Aflatoxins (AF) are toxic metabolites produced by two major fungi; Aspergillus (A.) flavus and A. parasiticus [13]. AFM1 is the type of aflatoxin that is present in milk and dairy products. Hydroxylation of AFB1 results in the formation of AFM1 which has been categorized as a class 1 human carcinogen [14]. The regulatory limits for AFM 1 vary in different countries; according to the regulations of the USA, levels of AFM1 in milk should not be higher than 500 ng/kg [15]. The ministry of health in Egypt established that milk should be free from AFM1 [16]. However, the European commission has stipulated that the maximum limit of AFM1 in liquid milk is 50 ng/kg, which is lower than the limit set by the Codex Alimentarius Commission [17] with 500 ng/kg [18].

In Egypt, Buffalo milk is produced at small-holder farms (unorganised) and mostly marketed without bactofugation, thermization or pasteurization treatment resulting in a significant decrease in the quality of raw market milk. Furthermore, there are few studies conducted on the quality of raw market Buffalo milk in the different regions of Egypt. Therefore, this study was undertaken to examine selected parameters of chemical and microbiological qualities as well as levels of AFM1 in raw milk samples taken at farmers’ markets of three different central governorates ( Cairo, Giza and Qualubya). The samples were collected from bulk tanks, in which the temperature of milk varied from 3 to 7 °C.
Considering the economic potential of buffalo milk and the growing awareness of Egyptian consumers regarding the importance of food safety, the results obtained will be useful in future studies dealing with the development of hygienic conditions in the production and marketing of buffalo milk.

**Material and Methods**

**Sampling:**
One hundred and twenty samples of raw buffalo milk were collected from markets at central locations of three different governorates (Cairo, Giza and Qualubya, 40 samples for each governorate) from November 2015 to February 2016 and stored at 4±1 °C until further analysis.

**Physico-chemical analyses:**
Raw milk samples were analysed for freezing point, total solids (T.S.), fat, protein and lactose using ultrasonic milk analyser (Lactoscan, Mik lotion Co., Nova Zagora, Bulgaria) in order to give us a quick analysis of the samples. Ash was measured according to Ling (1963) [19].

**Microbiological analyses:**
The pour plate technique with subsequent decimal dilutions in peptone saline (0.1 % neutral peptone and 0.9 % NaCl) as well as duplicate counting plates of appropriate dilutions were used for each sample. Standard plate count (32 °C /48 hours) was used to enumerate total bacterial count (TBC), MacConkey agar (37 °C /24 hours) was applied to enumerate coliform bacteria. Penicillin and pimaricin agar (PPA, 25 °C/48 hours) was used to enumerate psychrotrophic bacteria. All microbiological media were prepared according to the Oxoid manual [20], however, the PPA medium was purchased from Biolife Co., Milan, Italy.

**Determination of aflatoxin M1 levels:**
Twenty milliliters (20 ml) of liquid milk were centrifuged at 3500 xg / 7 °C. The fatty layer was removed and 100 μl of the skim milk was applied directly to the ELISA kit for AFM1 determination (R-Biopharm Co., RIDASCREEN® aflatoxin M1, Cat. No. R1121, Darmstadt, Germany).

**Statistical analysis:**
Microsoft Excel 2010 was used for data processing. Data are presented as mean ± standard deviation (SD) of two replicates for each sample. Results were subjected to one-way analysis of variance (ANOVA) through the general linear model (GLM) and multiple comparisons were performed using the Tukey test; the statistical significance was set at P < 0.05. All analyses were performed using SAS software version 9.4.

**Results and Discussion**

**Physical and chemical analyses:**
Results of the physical and chemical analyses of raw milk samples are presented in Table (1). The range, showing the minimum and maximum values, of the freezing point in raw market buffalo milk samples collected from central Cairo, Giza and Qualubya was -0.25 to -0.55, -0.39 to -0.55 and -0.25 to -0.55 °C respectively. The differences between the mean values of Giza and Qualubya were significant, but the decrease of mean values of the freezing point in all raw milk samples (average of 120 samples; -0.47 °C±0.07) of three different areas indicates that some of the raw milk samples were adulterated with water [5].

Data of the chemical composition (Table 1) of all samples were studied by variance analysis. A range of 4.18-7.20, 4.75-6.95 and 4.18-7.15% was found for fat, 2.85-4.93, 3.30-5.02 and 2.85-4.87 for protein, 2.55-4.87, 3.70-4.98 and 2.56-4.80 for lactose, 0.35-0.88, 0.49-0.89 and 0.37-0.84 for ash, 11.36-16.41, 12.18-16.44 and 11.38-16.41 for total solids contents in central of Cairo, Giza and Qualubya respectively. These results indicate that approx. 58% of the samples were not in accordance with Egyptian standards [7] for Buffalo milk (fat content is not less than 5.50 % and solids not fat (SNF) content is not less than 8.75 %), despite of the fact that the mean value or the average of all parameters were in accordance with the national standard as shown in the same Table. Our results are not in accordance with Najdenova et al. (2003) [21] and Supino et al. (2004) [22], because the average of fat content in buffalo milk was 7.5 %. Also, the mean concentrations from one hundred buffalo milk samples were 7.59±1.31 % for fat, 4.86±0.44 % for crude protein, 0.85±0.05 % for ash and 18.44±1.56 % for total solids [23]. There are several factors such as type of breed, lactation period, forage, feeding system, seasonal changes, milking frequency and milking methods which can impact the physicochemical parameters as described by Suman et al. (1998) [24].

**Microbiological analyses:**
The total bacteria counts (TBC), coliform, spore forming bacteria and psychrotrophic are presented in Table (2). A difference in the mean of the tested bacterial groups in all milk samples was noticed, which might be due to differences in the sanitation process of the different farmers’ markets. Also, 36 % of samples had higher TBC than 6 Log CFU/mL. However, the average of TBC for all samples (5.37±0.78) was in accor-

<table>
<thead>
<tr>
<th>Area</th>
<th>Parameter</th>
<th>FP °C</th>
<th>Fat</th>
<th>Protein</th>
<th>Lactose</th>
<th>Ash</th>
<th>Total Solids</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Central Cairo (n=40)</strong></td>
<td>Range</td>
<td>0.25-0.55</td>
<td>4.18-7.20</td>
<td>2.85-4.93</td>
<td>2.55-4.87</td>
<td>0.35-0.88</td>
<td>11.36-16.41</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>0.46±0.01</td>
<td>5.83±0.90</td>
<td>4.10±0.59</td>
<td>4.21±0.61</td>
<td>0.67±0.15</td>
<td>14.54±1.56</td>
</tr>
<tr>
<td><strong>Central Giza (n=40)</strong></td>
<td>Range</td>
<td>0.39-0.55</td>
<td>4.75-6.95</td>
<td>3.30-5.02</td>
<td>3.70-4.98</td>
<td>0.49-0.89</td>
<td>12.82-16.44</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>0.49±0.05</td>
<td>6.11±0.61</td>
<td>4.28±0.44</td>
<td>4.39±0.38</td>
<td>0.73±0.11</td>
<td>14.96±1.10</td>
</tr>
<tr>
<td><strong>Central Qualubya (n=40)</strong></td>
<td>Range</td>
<td>0.25-0.55</td>
<td>4.18-7.15</td>
<td>2.85-4.87</td>
<td>2.56-4.80</td>
<td>0.37-0.84</td>
<td>11.38-16.41</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>0.45±0.08</td>
<td>5.74±0.82</td>
<td>4.01±0.53</td>
<td>4.16±0.56</td>
<td>0.66±0.14</td>
<td>14.29±1.43</td>
</tr>
<tr>
<td><strong>Average (n=120)</strong></td>
<td>Range</td>
<td>0.25-0.55</td>
<td>4.18-7.20</td>
<td>2.85-5.02</td>
<td>2.55-4.98</td>
<td>0.35-0.89</td>
<td>11.36-16.44</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>0.47±0.07</td>
<td>5.89±0.79</td>
<td>4.13±0.53</td>
<td>4.25±0.53</td>
<td>0.69±0.14</td>
<td>14.59±1.39</td>
</tr>
</tbody>
</table>

*Data are expressed as the mean ± standard deviation (SD), samples were analysed in duplicate. Means with different superscript letters differ significantly (p<0.05).*

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**Table 1: Physical and chemical properties of samples collected from local markets of three different central governorates in Egypt**

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dance with the Egyptian standards [7] for raw buffalo milk, being 5.70 Log CFU/mL. Results of TBC in raw buffalo milk samples indicate that there are inadequate sanitary conditions during production and marketing of raw buffalo milk. In Italy, the TBC of raw buffalo milk was 5.23 Log CFU/mL [22] and it was 5.59 Log CFU/mL in China [23]. However, the TBC average of buffalo milk samples in the Alexandria Governorate was 6.70 Log CFU/mL [25] and in the Menoufia Governorate, Egypt, 7.60 Log CFU/mL [26].

The occurrence of the Coliform group and E. coli in milk indicates poor hygiene or fecal contamination. Other enteric pathogens may also originate from the external surface during manual milking [27]. Results in Table (2) show that the levels of the coliform group in the three different governorates were higher than the acceptable levels in both the Egyptian standards [7] and the EU specification [28] where the average coliform count was 3.77 Log CFU/mL. Furthermore, 46% of the samples had a coliform count of more than 4 Log CFU/mL. Our results are not in accordance with results obtained by Han et al. (2007) [23] who reported that the average coliform count in buffalo milk samples was 2.42 Log CFU/mL. Results obtained by Ombarak and ElBagory (2015) [26] showed higher levels of coliform bacteria (with ca. 5.90 Log CFU/mL) in raw buffalo milk samples collected from local markets in Menoufia.

Spore forming bacteria (such as Bacillus sp. and Clostridium sp.) derive from the farm environment. They can well survive during the pasteurization of milk [29] and grow during refrigerated storage [30] causing milk and its products to spoil and limiting their shelf life [31]. Although there are some regional, seasonal and methodological differences, the average counts of aerobic spore forming bacteria (mostly predominance Bacillus licheniformis) in raw milk are 10 to 10^2 CFU mL^-1 [32-34]. The range of aerobic spore forming bacteria between the different tested regions was relatively the same (Table 2). The relative stability of the spore forming bacteria counts in the three different regions might be due to the similarity of milk chain productions from farmers located in Giza and Qualubya to retailers. Here we also analysed samples of bulk milk (collected from different sources) at farmers’ markets and were unable to find significant variations in the viable count of spore forming bacteria between tested regions. Results of the analysis of one hundred and twelve buffalo milk samples in China showed that the average of bacterial endospores was 2.31 Log CFU mL^-1 [23]. In the end improving management practices and forage quality could be a useful solution in decreasing the contamination of milk with spore forming bacteria [35].

Psychrotrophic bacteria are defined as those growing at 7 °C. They represent the dominant microflora during the cold storage period and produce heat resistant lipases and proteases causing different dairy products to spoil [36]. Our data in Table (2) show that there was no big variation in the range of psychrotrophic bacteria between central Cairo (2.14 to 3.51), Giza (2.14-3.37) and Qualubya (2.12-3.48). The average psychrotrophic bacteria count for all samples was 2.84 Log CFU/mL, indicating the absence of proper standardization of cold storage systems during production and marketing or raw buffalo milk. Our results are slightly in accordance with Cempírková (2002) [37] and Cempírková et al. (2009) [38] who reported the average of psychotrophic bacteria in samples of cow milk at 3.00 and 3.41 Log CFU/mL. In another study, the range of psychrotrophic bacteria in goat milk samples of twelve farms was 2.9 to 5.0 Log CFU/mL [39].

Levels of AFM1 in raw milk samples:

Results (Figure 1) show that levels of AFM 1 in central Qualubya (143.58 ng/L) are significantly (P < 0.05) higher than in central Cairo (57.26 ng/L) and central Giza (29.68 ng/L). This result may be due to the similarity of milk chain productions from farmers located in Giza and Qualubya to retailers. Here we also analysed samples of bulk milk (collected from different sources) at farmers’ markets and were unable to find significant variations in the viable count of spore forming bacteria between tested regions. Results of the analysis of one hundred and twelve buffalo milk samples in China showed that the average of bacterial endospores was 2.31 Log CFU mL^-1 [23]. In the end improving management practices and forage quality could be a useful solution in decreasing the contamination of milk with spore forming bacteria [35].

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**Table 2: Microbiological properties of samples collected from local markets of three different central governorates in Egypt**

<table>
<thead>
<tr>
<th>Area</th>
<th>Parameter</th>
<th>Total bacterial count</th>
<th>Coliform</th>
<th>Spore forming bacteria</th>
<th>Psychrotrophic bacteria</th>
</tr>
</thead>
<tbody>
<tr>
<td>Central Cairo (n=40)</td>
<td>Range</td>
<td>4.15-6.59</td>
<td>2.15-4.68</td>
<td>2.25-3.43</td>
<td>2.14-3.51</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>5.41±0.83 ab</td>
<td>3.85±0.86</td>
<td>2.81±0.36</td>
<td>2.88±0.40</td>
</tr>
<tr>
<td>Central Giza (n=40)</td>
<td>Range</td>
<td>4.21-6.35</td>
<td>2.10-4.68</td>
<td>2.25-3.45</td>
<td>2.14-3.37</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>5.22±1.00 b</td>
<td>3.54±0.91</td>
<td>2.71±0.33</td>
<td>2.74±0.35</td>
</tr>
<tr>
<td>Central Qualubya (n=40)</td>
<td>Range</td>
<td>4.15-6.59</td>
<td>2.15-4.68</td>
<td>2.25-3.43</td>
<td>2.12-3.48</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>5.50±0.80 a</td>
<td>3.92±0.88</td>
<td>2.84±0.39</td>
<td>2.89±0.41</td>
</tr>
<tr>
<td>Average (n=120)</td>
<td>Range</td>
<td>4.15-6.59</td>
<td>2.10-4.68</td>
<td>2.25-3.45</td>
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<td></td>
<td>Mean</td>
<td>5.37±0.78</td>
<td>3.77±0.90</td>
<td>2.78±0.36</td>
<td>2.84±0.39</td>
</tr>
</tbody>
</table>

Data are expressed as the mean ± standard deviation (SD), samples were analysed in duplicate. Means with different superscript letters differ significantly (p<0.05).

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**Figure 1. Aflatoxin M1 content of samples collected from local markets of three different central governorates in Egypt.** The box and whiskers plot displays the entire range of values, IQR (interquartile range; 25% - 75%, median(-), mean(x)), samples were analysed in duplicate. Means with different superscript letters within row differ significantly (p<0.05).
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compared to the other two governorates. The median concentration of AFM 1 in all samples was 44.31 ng/L. This level of AFM1 is near to the permissible limit of 50 ng/L set by EU regulations [40] and higher than the limit of zero ng/L set by Egyptian regulations [16]. Finally, concentrations of AFM 1 in 27 % of the milk samples were less than the limit of detection (5 ng/L). In the Alexandria governorate, the mean level of AFM1 in fifty samples of raw cow milk was 49.74 ng/L [41]. However, the average level of AFM1 in thirty samples of raw buffalo milk in the Sohag and Assiut governorates was 64.49 and 130.60 ng/L respectively [42].

Conclusion
We analysed one hundred and twenty samples of market buffalo milk for selected physico-chemical and microbiological parameters as well as levels of AFM1. Our results indicate that there is a serious hygiene problems under the Egyptian regulations. Therefore, we recommend that good hygienic practices and regulations, such as improving farm management practices and implementing the ISO 22000 food safety management system including HACCP to facilitate the production of high quality and safe buffalo milk. Indeed, extensive research is still needed regarding the characterization of predominant spore forming bacteria and other microorganisms causing spoilage in Egyptian buffalo milk.

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References


